

- R. ZAMUDIO, J. BOSCH, S. LOTTERS, E. WOMBWELL, T. W. J. GARNER, A. A. CUNNINGHAM, A. SPITZEN-VAN DER SLUIJS, S. SALVIDIO, R. DUCATELLE, K. NISHIKAWA, T. T. NGUYEN, J. E. KOLBY, I. VAN BOCXLAER, F. BOSSUYT, AND F. PASMANS. 2014. Recent introduction of a chytrid fungus endangers Western Palearctic salamanders. *Science* 346:630–631.
- MITCHELL, J. C. AND K. A. BUHLMANN, K. A. 1999. Amphibians and reptiles of the Shenandoah Valley sinkhole pond system in Virginia. *Banisteria* 13:129–142.
- MOFFITT, D., L. A. WILLIAMS, A. HASTINGS, M. W. PUGH, M. M. GANGLOFF, AND L. SIEFFERMAN. 2015. Low prevalence of the amphibian pathogen *Batrachochytrium dendrobatidis* in the Southern Appalachian Mountains. *Herpetol. Conserv. Biol.* 10:123–136.
- O'HANLON, S. J., A. RIEUX, R. A. FARRER, G. M. ROSA, B. WALDMAN, A. BATAILLE, T. A. KOSCH, K. A. MURRAY, B. BRANKOVICS, M. FUMAGALLI AND M. D. MARTIN. 2018. Recent Asian origin of chytrid fungi causing global amphibian declines. *Science* 360:621–627.
- OLSON, D. H., K. L. RONNENBERG, C. K. GLIDDEN, K. R. CHRISTIANSEN, AND A. R. BLAUSTEIN. 2021. Global patterns of the fungal pathogen *Batrachochytrium dendrobatidis* support conservation urgency. *Front. Vet. Sci.* 8:685877.
- RICHGELS, K. L., R. E. RUSSELL, M. J. ADAMS, C. L. WHITE, AND E. H. C. GRANT. 2016. Spatial variation in risk and consequence of *Batrachochytrium salamandrivorans* introduction in the USA. *Roy. Soc. Open Sci.* 3:150616.
- SCHEELE, B. C., F. PASMANS, L. F. SKERRATT, L. BERGER, A. N. MARTEL, W. BEUKEMA, A. A. ACEVEDO, P. A. BURROWS, T. CARVALHO, A. CATENAZZI, I. DE LA RIVA, M. C. FISHER, S. V. FLECHAS, C. N. FOSTER, P. FRIAS-ALVAREZ, T. W. J. GARNER, B. GRATWICKE, J. M. GUAYASAMIN, M. HIRSCHFELD, J. E. KOLBY, T. A. KOSCH, E. LA MARCA, D. B. LINDEMAYER, K. R. LIPS, A. V. LONGO, R. MANEYRO, C. A. McDONALD, J. MENDELSON III, P. PALACIOS-RODRIGUEZ, G. PARRA-OLEA, C. L. RICHARDS-ZAWACKI, M. RODEL, S. M. ROVITO, C. SOTO-AZAT, L. F. TOLEDO, J. VOYLES, C. WELDON, S. M. WHITFIELD, M. WILKINSON, K. R. ZAMUDIO, AND S. CANESSA. 2019. Amphibian fungal panzotic causes catastrophic and ongoing loss of biodiversity. *Science* 363:1459–1463.
- WADDLE, J. H., D. A. GREAR, B. A. MOSHER, E. H. C. GRANT, M. J. ADAMS, A. R. BACKLIN, W. J. BARICHIVICH, A. B. BRAND, G. M. BUCCIARELLI, D. L. CALHOUN, T. CHESTNUT, J. M. DAVENPORT, A. E. DIETRICH, R. N. FISHER, B. M. GLORIOSO, B. J. HALSTEAD, M. P. HAYES, R. K. HONEYCUTT, B. R. HOSACK, P. M. KLEEMAN, J. A. LEMOS-ESPINAL, J. M. LORCH, B. MCCREARY, E. MUTHS, C. A. PEARL, K. L. D. RICHGELS, C. W. ROBINSON, M. F. ROTH, J. C. ROWE, W. SADINSKI, B. H. SIGAFUS, I. STASIAK, S. SWEET, S. C. WELLS, G. J. WATKINS-COLWELL, C. LEANN WHITE, L. A. WILLIAMS, AND M. E. WINZELER. 2020. *Batrachochytrium salamandrivorans* (*Bsal*) not detected in an intensive survey of wild North American amphibians. *Sci. Rept.* 10:13012.
- WEINSTEIN, S. B. 2009. An aquatic disease on a terrestrial salamander: individual and population level effects of the amphibian chytrid fungus, *Batrachochytrium dendrobatidis*, on *Batrachoseps attenuatus* (Plethodontidae). *Copeia* 2009:653–660.
- YAP, T. A., N. T. NGUYEN, M. SERR, A. SHEPACK, AND V. T. VREDENBURG. 2017. *Batrachochytrium salamandrivorans* and the risk of a second amphibian pandemic. *EcoHealth* 14:851–864.

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## First Detection of Apparent Ophidiomycosis in the Mole Kingsnake (*Lampropeltis rhombomaculata*) in North Carolina, USA

Understanding the full host and geographic extent of ophidiomycosis caused by the fungal pathogen *Ophidiomyces ophidiicola* is an emerging priority (Haynes and Allender 2021), especially given recent molecular support for its introduction

**EMILY OVEN**

**ALEXANDRIA NELSON**

**ZACHARY GAJEWSKI**

Department of Applied Ecology, North Carolina State University,  
 Raleigh, North Carolina 27607, USA

**CHRISTOPHER S. DEPERNO**

Department of Forestry and Environmental Resources, North Carolina  
 State University, Raleigh, North Carolina 27607, USA

**MATTHEW C. ALLENDER**

**LAURA ADAMOVICZ**

Wildlife Epidemiology Lab, Veterinary Diagnostic Lab, University of  
 Illinois Urbana-Champaign, Urbana, Illinois 61802, USA

**SKYLAR R. HOPKINS\***

Department of Applied Ecology, North Carolina State University,  
 Raleigh, North Carolina 27607, USA

\*Corresponding author; e-mail: skylar\_hopkins@ncsu.edu

to the United States (Ladner et al. 2022). Although the Mole Kingsnake, *Lampropeltis rhombomaculata*, is widespread in the southeastern USA, it has rarely been surveyed for pathogens or other metrics of health due to its cryptic and fossorial life history. In one prior case, a single individual was investigated for ophidiomycosis (formally known as snake fungal disease) by the Wildlife Epidemiology Laboratory at the University of Illinois Urbana-Champaign after exposure to another snake infected with *O. ophidiicola*. That individual never developed clinical signs or tested positive for *O. ophidiicola* DNA. There have been no prior surveys for ophidiomycosis in wild *L. rhombomaculata*, but ophidiomycosis has been reported from six other *Lampropeltis* species (Haynes and Allender 2021), suggesting that *L. rhombomaculata* is likely to be susceptible to the disease. Here, we report the first case of suspected ophidiomycosis in a wild *L. rhombomaculata*.

At 0855 h on 13 August 2022, we discovered an adult *L. rhombomaculata* (81.5 cm SVL, 12.5 cm tail length) under a partially decayed log while surveying terrestrial colubrids at G. W. Hill Forest, Durham County, North Carolina, USA (36.20083°N, 78.88525°W; WGS 84). At the time of capture, the ambient air temperature was 21.8°C and humidity was 74.8%, and under the





FIG. 1. Clinical signs of ophidiomycosis on a *Lampropeltis rhombomaculata* from Durham County, North Carolina, USA: A) general signs of ecdysis; B) dry crusts visible on the dorsal body. For both photographs, the snake was left unaltered, but we slightly blurred the background.

log, the temperature was 19.7°C and the soil moisture was 10%. The snake was beginning ecdysis, with opaque eye scales and pale body coloring (Figure 1A). Also, the snake presented with clinical signs of ophidiomycosis in the form of several crusts on the dorsal body, measuring approximately 3 cm by 1 cm in size (Figure 1B). We collected two skin swabs (one dorsal, one ventral) by passing the swab along the entire length of the body from snout to tail tip five times, including the lesions.

Both swabs later tested positive for *O. ophidiicola* DNA using a previously published qPCR assay (Allender et al. 2015). Because this snake had consistent clinical signs of ophidiomycosis and a positive qPCR test, it is classified as a case of “apparent ophidiomycosis” (“confirmed ophidiomycosis” requires a skin biopsy and histological analysis; Baker et al. 2019). Additional surveillance and monitoring are warranted to better understand the scope of *L. rhombomaculata* infection and the significance of infection for individuals, populations, and communities.

#### LITERATURE CITED

- ALLENDER, M. C., D. BUNICK, E. DZHAMAN, L. BURRUS, AND C. W. MADDOX. 2015. Development and use of a real-time polymerase chain reaction assay for the detection of *Ophidiomyces ophidiicola* in snakes. *J. Vet. Diag. Invest.* 27:217–220.
- BAKER, S. J., E. HAYNES, M. GRAMHOFER, K. STANFORD, S. BAILEY, M. CHRISTMAN, K. CONLEY, S. FRASCA, JR., R. J. OSSIBOFF, D. LOBATO, AND M. C. ALLENDER. 2019. Case definition and diagnostic testing for snake fungal disease. *Herpetol. Rev.* 50:279–285.
- HAYNES, E. K. AND M. C. ALLENDER. 2021. History, epidemiology, and pathogenesis of ophidiomycosis: A review. *Herpetol. Rev.* 52:521–536.
- LADNER, J. T., J. M. PALMER, C. L. ETTINGER, J. E. STAJICH, T. M. FARRELL, B. M. GLORIOSO, B. LAWSON, S. J. PRICE, A. G. STENGLE, D. A. GREAR, AND J. M. LORCH. 2022. The population genetics of the causative agent of snake fungal disease indicate recent introductions to the USA. *PLoS Biol.* 20:e3001676.